

Preparation of samples for sci- or 10x-ATAC-seq

Primary or cultured cells

- 1) Resuspend cells in 0.5 mL of media containing 10% DMSO (Input: sci-ATAC-seq: $\geq 1M$ cells; 10x-scATAC-seq: $>250k$ cells; 10x-sc/snRNA + sci/10x-ATAC-seq (from same sample): $\geq 500k$ cells)

Note: The critical factor is that the resuspension medium contains 10% DMSO or equivalent cryoprotectant. Cells frozen down without cryoprotectant are not amenable to ATAC-seq. In our experience, the other media components used at this stage are not critical.

- 2) Slow freeze in isopropanol bath or similar, at $-80^{\circ}C$.

Note: In our experience, it is also fine to flash freeze the cells directly in liquid nitrogen. In some cases, we find that slow freezing can lead to slightly better signal-to-noise ratio.

After freezing, cells can be stored for months at $-80^{\circ}C$.

Tissues

Tissues should be flash-frozen in liquid nitrogen immediately upon resection. If unable to freeze immediately, we recommend storing in ice-cold PBS in the interim. Tissues should be unfixed. (Input: sci-ATAC-seq: $\geq 500k$ nuclei; 10X-ATAC-seq: $\geq 250k$ nuclei; 10x-sc/snRNA + sci/10x-ATAC-seq (from same sample): $\geq 500k$ nuclei)

General Notes:

- 1) **Important:** Before preparing a large set of samples for sci or 10x-ATAC-seq, we strongly recommend submitting 1-2 test samples of the same type and using the same preparation method as the larger batch of samples to be assayed.
- 2) If you cannot fulfill the sample input requirements, please contact epigenome@ucsd.edu we will find a custom solution for you.
- 3) We prefer that samples be submitted in standard 1.5 mL microfuge tubes or 2 mL cryotubes. If you require another format, please contact epigenome@ucsd.edu to make arrangements prior to sample submission.