

Diversity at the 5-Carbon Position of Sialic Acids

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This chapter discusses our recent work on two modifications at the 5-position of sialic acids. Earlier studies suggested that humans may not express *N*-glycolyl-neuraminic acid (Neu5Gc), a hydroxylated form of *N*-acetyl-neuraminic acid (Neu5Ac). We found that while traces of Neu5Gc are detectable in human body fluids and tissues, it is a major component in all the great apes which are our closest evolutionary relatives. This correlates with a human deficiency of the hydroxylase that converts CMP-Neu5Ac to CMP-Neu5Gc. While the chimpanzee hydroxylase gene is similar to the murine homologue, the human gene contains a 92 bp deletion, resulting in a frame-shift mutation. Thus, the lineage leading to modern humans suffered this mutation sometime after our common ancestor with the chimpanzee and bonobo, potentially affecting recognition of cell surface sialic acids by many endogenous and exogenous sialic acid binding lectins. The expression of Neu5Gc previously reported in human fetuses and tumors, as well as the traces detected in normal humans must be mediated by an alternate pathway. In unrelated work, we found that monoclonal antibody SGR37 detects de-*N*-acetylation of the 5-*N*-acetyl group of the disialoanglioside G_{D3} (giving DeNAc- G_{D3}). While this antigen is expressed at low levels in a few normal human cells, it accumulates at high levels in many melanomas and in some lymphomas, but not in carcinomas. Isotype-matched antibodies against G_{D3} , 9-OAc- G_{D3} and DeNAc- G_{D3} showed that while the first two are predominantly on the cell surface and partly in lysosomal compartments, DeNAc- G_{D3} has a diffuse intracellular location, which appears to result via endocytosis from the cell surface. Rounding up of melanoma cells during growth is associated with relocation of the internal pool to the cell surface. Thus, a minor modification of a tumor-associated ganglioside can markedly affect subcellular localization and trafficking. There is also potential for use of the antibody in diagnostic or therapeutic approaches to these tumors.

Structural Diversity at the 5-Carbon Position of Sialic Acids

The sialic acids are a family of 9-carbon monosaccharides that occur in animals of the Deuterostome lineage (vertebrates and "higher" invertebrates) (1). They are typically located at the outermost ends of *N*-glycans, *O*-glycans and Glycosphingolipids (they can also occasionally cap the side chain of GPI-anchors). Sialic acids are subject to a remarkable variety of modifications (1-3) which are the subjects of several presentations in this volume. This chapter focuses exclusively on structural variations that can occur at the 5-carbon position of these molecules. Most commonly, there is an *N*-acetyl group at this position (giving *N*-acetyl-neuraminic acid or Neu5Ac). Less commonly there is hydroxyl group (giving keto-deoxy-nonulosonic acid, KDN)

(4). The 5-*N*-acetyl group can also be hydroxylated. The addition of this single oxygen atom gives *N*-neuraminic acid (Neu5Gc) (1-3). The hydroxylated glycolyl group can even be further substituted with an acetyl group (3). Rarely, the 5-*N*-acetyl group is deleted, giving neuraminic acid (Neu)(5-9). This free hydroxyl group at the 5-position can in turn, react covalently with a carboxylate group at the 1-carbon position, giving a lactam (10). Unsaturated and dehydro-forms of these sialic acids are also known to exist (1). This chapter focuses on our recent work on the natural occurrence of two of these sialic acids: *N*-glycolyl-neuraminic acid (Neu5Gc) (11,12) and neuraminic acid (Neu5Ac). **Human deficiency of *N*-glycolyl-neuraminic acid** Neu5Gc differs from Neu5Ac by a single oxygen,

added by the action of a specific hydroxylase that can convert the nucleotide sugar donor CMP-Neu5Ac to CMP-Neu5Gc (13–23). The expression of Neu5Gc is known to be widespread in mammalian cells - in fact, it seems to be the major sialic acid in some species. It also shows tissue-specific and developmentally-regulated expression in a variety of systems (1,2,24–26). This modification of the parent molecule Neu5Ac can affect interactions involving many known endogenous and exogenous receptors for sialic acids such as the siglecs and virus hemagglutinins (27–30).

Early pioneers in the sialic acid field noted that in contrast to many other animals, human tissues did not have easily detectable Neu5Gc (1). In other work, it was shown that the immune response of humans to the infusion of animal serum was at least partly directed against the Neu5Gc epitope present in the infused serum (31,32). Some humans with cancer or certain other diseases can also spontaneously express antibodies directed against Neu5Gc (31,33–35). Human fetal tissues (36) and certain human tumors (35–37) were also reported to contain small amounts of Neu5Gc. However, one study did not support the presence of Neu5Gc in human tumor cells (38). Also, most of the above observations were made using monospecific antibodies and/or TLC analysis. Analysis by TLC can be limited in sensitivity and specificity, and antibodies against carbohydrates are known to cross-react with other related structures. We therefore decided to re-explore the issue of Neu5Gc expression in humans, and in their evolutionary cousins, the great apes.

Great apes express N-glycolyl-neuraminic acid at levels comparable to that of other animals. All humans living today belong to a single species with limited genetic diversity (39). Darwin and Huxley correctly deduced that humans are most closely related to the African great apes (40,41), and this prediction has been amply confirmed by a variety of methods (42–51). From the strictly genetic point of view humans should be classified simply as specialized members of the great ape family, since we share an almost 99% genomic DNA sequence identity with *Pan troglodytes* (the chimpanzee) and *Pan paniscus* (the bonobo) which, in turn, have a lesser genetic identity with *Gorilla gorilla* (the gorilla) and *Pongo pygmaeus* (the orangutan) (49–51). Thus, the obvious morphological and functional differences between humans and these great apes should be based on relatively few changes in gene expression or function. How-

ever, despite a large amount of comparison over the years, there have been no clearcut differences in the expression of gene products between humans and great apes.

We found that while Neu5Gc is ubiquitous in all plasma proteins and erythrocytes, it is also present in all four great apes, as well as in mice (11). With our collaborator Elaine M. Schmitt we found that this marked difference in Neu5Gc expression was seen amongst cultured lymphoblastoid cell lines from humans and great apes, as well as in a variety of tissues compared between humans and chimpanzees (11). In the cerebral cortex and the cerebrospinal fluid, the above findings, CMP-sialic acid I was detected in chimpanzee cells, but not in humans (11). However, we did notice traces of Neu5Gc in some normal human cells (11). Confirmation of this finding by mass spectrometry is under way. As mentioned above, Neu5Gc has also been reported by others in human tissues (13, 15, 16, 18, 19, 21–23). The regulation of Neu5Gc activity is very complex, and involves many factors (13, 15, 16, 18, 19, 21–23). It has also appeared that the activity of this hydroxylase is suppressed in adult humans, rather than in other great apes.

A mutation in human CMP-sialic acid hydroxylase occurred after our last common ancestor. To explore this issue, we identified and sequenced base sequences homologous to the mouse hydroxylase cDNA, and used them to identify cDNAs for the chimpanzee and human. DNA Sequencing indicated a 92 bp deletion of the human cDNA. In contrast, the chimpanzee cDNA sequence is functionally intact and identical to that of the mouse. The deletion results in a frame shift, which would cause a truncation of the coding sequence. Since the chimpanzee sequences coding for the putative Reissinger region of the murine hydroxylase predicted molecule is highly unlikely to have any activity. This deletion in the cDNA is reported independently by Irie, *et al.* (52) prior to our publication. However, it

isolate the complete 5' region of the human cDNA that includes the primary initiator methionine. Thus their 5' truncated cDNA gave a different protein product in an *in vitro* translation reaction, leading to the assumption that a methionine codon downstream of the 92 bp deletion is the primary initiator for translation of a truncated protein representing >80% of the carboxyl end of the hydroxylase. They went on to show that such a truncated protein had no residual hydroxylase activity when expressed in COS cells. However, we find an intact primary initiator methionine codon corresponding to that of the mouse and chimpanzee protein, and therefore predict a much shorter truncated polypeptide involving the amino terminus of the protein. Thus, it remains to be shown that the protein product studied *in vitro* by Irie et al. actually *exists in vivo*. Regardless, both groups agree that humans are genetically deficient in the activity of the hydroxylase.

Genomic PCR analysis indicated that the corresponding 92 bp region is present in total genomic DNA of all the African great apes, indicating that this region of the human genome is either deleted or markedly altered in sequence (12). We also found that human genomic BAC clones spanning the hydroxylase gene failed to hybridize with a chimpanzee cDNA probe corresponding to the 92 bp human cDNA deletion (12). Irie, *et al.* (53) went further and actually sequenced the entire human genomic region in question, proving that the 92-bp loss represents an exon deletion corresponding to exon 6 of the mouse hydroxylase gene. Chromosomal localization studies by our collaborator David Nelson showed that the hydroxylase gene is located at 6p22-p23 (12). This is not close to the site of any of the major chromosomal rearrangements known to have occurred in hominoid evolution (54).

The 92 bp deletion is found in diverse human populations. Our studies of Neu5Gc deficiency in humans included a survey of more than 70 individuals from many ethnic backgrounds (11). To confirm that the hydroxylase mutation is not simply a human polymorphism, our collaborator S. Warren carried out a similar PCR analyses on DNA samples from 18 "Caucasians", 4 African-Americans, 4 !Kung bushmen, 4 Khwe pigmies and 6 Japanese. All these samples failed to give the 92 bp PCR product, while all control samples from chimpanzees and bonobos showed the product (12). Thus, the human CMP-Neu5Ac hydroxylase

gene seems to have undergone this inactivation after the time of the last common ancestor with and chimpanzee, but before the common origin of modern humans.

What are the consequences for human evolution? The change from Neu5Ac to Neu5Gc could theoretically affect several of the known endogenous and exogenous receptors for sialic acids (27, 28, 30). The interactions of sialic acids with pathogens such as Influenza A and B viruses (*E. coli* K99 (58–61) are known to be affected to some extent. There are many other microbes that utilize sialic acids as specific binding sites on mammalian cells. For example, several major pathogens such as *Helicobacter pylori* and *Modiolum falciparum* (62–65). In most such cases, sequences of having Neu5Gc or Neu5Ac have not been studied. Sialic acids are also ligands for several vertebrate lectins (27, 28, 30, 63, 66–68). For example, Siglecs such as sialoadhesin on macrophages, and associated glycoprotein on neuronal axon myelin show a lack of binding to Neu5Gc (27). Thus, the demands causing loss of Neu5Gc expression could influence a variety of normal and abnormal processes. Interestingly, even in animals that express large amounts of Neu5Gc in other tissues, the level in the brain is extremely low (69–71). The reason for this is unknown, but it is tempting to speculate that humans have gained an advantage in brain development and/or function by eliminating the last traces of Neu5Gc in the brain, while "saccharides" are present throughout the rest of the body.

Finally, we have also isolated alternative human cDNAs with various deletions and insertions. Although we did not conduct an extensive search in a chimpanzee, we encountered only one such anomaly among chimpanzee clones that we obtained. Thus, we speculate that the many abnormalities and variations of the 3' end of the human hydroxylase are indicative of a gene that is non-functional, and is undergoing a random degeneration as a pseudogene. Regardless, if it were possible to determine when exactly the loss of Neu5Gc expression occurred in relation to hominoid evolution, this could help to resolve a controversial question of the origin of modern humans as well as make predictions regarding the functional consequences of Neu5Gc loss. Unfortunately, autosomal genes are extremely difficult to recover from fossils >30,000 years old.

old (73). We are currently extracting and studying the sialic acids themselves from fossil samples.

What about the Traces of Neu5Gc Found in Human Tissues, Fetuses and Tumors?

Despite the finding of an inactivating mutation in the human hydroxylase gene, human fetal tissues (36) and certain human tumors (36, 37,74) have been reported to contain small amounts of Neu5Gc. In keeping with this, some humans with malignancies or with certain inflammatory or infectious diseases are known to spontaneously develop antibodies against Neu5Gc (31, 33, 34). As previously mentioned, we have also found small traces of a HPLC peak corresponding to Neu5Gc in some human tissues (11). Taken together, these data suggest that the ability to express Neu5Gc may not be completely lost in humans. We need to explore if this represents the upregulation of another minor pathway for the synthesis of Neu5Gc, such as the use of the donor glycolyl-CoA (75,76), or the incorporation of Neu5Gc derived from dietary sources (77).

De-*N*-Acetyl-Gangliosides in Humans as Novel Tumor Antigens

Gangliosides are sialic acid-containing glycosphingolipids that are typically concentrated on the outer leaflet of the plasma membrane of vertebrate cells (78,79). They contribute to the physical properties of the membrane (80), can act as receptors for bacterial and viral adhesins (62), and can interact with and modulate the functions of growth factors and extracellular matrix receptors (81,82). Gangliosides are also thought to be potential ligands for endogenous animal lectins such as selectins (83, 84) and the siglecs (29, 30). The common monosialoganglioside G_{M3} (Sia α 2-3LacCer) can be modified by the ST8Sia I sialyltransferase(85,86) to give G_{D3} (Sia α 2-8Sia α 2-3LacCer), which is a known marker for cell activation and transformation (87-90). Such ganglioside antigens are also major targets for the immunotherapy of cancer (91-98).

The monoclonal antibodies (Mabs) SGR37 and SMR36 were raised by our collaborator Tadashi Tai, using chemically synthesized de-*N*-acetyl- G_{D3} as an immunogen. We had previously reported reactivity of these antibodies with human melanoma cell lines such as Melur and M21 (8). More recently, we compared the distribution of the SGR37

antigen (de-*N*-acetyl- G_{D3}) with that of G_{D3} in normal human tissues and in hu. SGR37 is an IgG₃ antibody that was highly specific manner with de-*N*-acetyl recognition a non-acetylated amino acid residue, as well as the intact sialic acid unit. As expected for glyco section reactivity was markedly diminished in organic solvents (*e.g.* methanol) or by affinity embedding process(99). Even fixation caused some loss of the SGR37 antigen. Unlike other glycolipid antigens, the way to circumvent this is to avoid using organic solvents, and instead use fixation with aldehydes. While this approach worked with mAb R24 (anti- G_{D3}), SGR37 reactivity was diminished at concentrations of paraformaldehyde fixation (100). This is probably because the free amino group, a major determinant of the SGR37 antigenicity, is modified by paraformaldehyde. In addition, similar problems were noted with other glycolipid antigens, and we therefore turned to the use of fresh unfixed tissue sections (100), which, while more difficult to process, showed consistent reactivity with the SGR37 antibody.

The SGR37 antigen is an uncommon structure in normal human

tissues. In a survey of both R24 and SGR37 antigenicity in normal human tissues using unfixed tissue sections, R24 antigen positivity was seen in melanocytes, blood vessels, pancreatic medullary cells, marginal zone lymphocytes in the spleen, interfollicular zone lymphocytes in the testis, and smooth muscle cells. SGR37 antigenicity was not seen as well with prior reports (90,91). In contrast, reactivity to SGR37 was found in melanocytes and in blood vessels of some tissues, and in the pia mater and white matter of the brain. Cells that were clearly negative for the SGR37 antigen were a few non-nuclear cells seen in the skin and colon.

Can we be certain that the antigens in tissue sections are indeed gangliosides? We have known to be highly specific for de-*N*-acetyl-ganglioside targets. The extractability of the

solvents or paraffin embedding (see above) suggests that the natural epitope is on a lipid, and the sensitivity of the SGR37 antigen to aldehydes (see above) fits with the requirement of this antibody for a free amino group. We also found that strong periodate oxidation (46 mM, neutral pH) abolished both SGR37 and R24 reactivity, indicating that the epitopes in the tissues for both antibodies are carbohydrate-dependent. Overall, the sensitivity to organic solvents and aldehydes as well as the abolition of reactivity by strong periodate reassures us that the molecule being detected by SGR37 in the tissues is either de-*N*-acetyl G_{D3} , or some closely related structural analogue. Thus, we can conclude that MAb SGR37, which was originally raised against a synthetic ganglioside recognizes a naturally occurring ganglioside antigen in a few normal human cell types. Since many R24 positive areas did not also stain for SGR37, it appears that GD3 expression is not always accompanied by de-*N*-acetylation.

The SGR37 antigen accumulates in most melanomas and some lymphomas, but not in carcinomas. Many clinical samples of human melanomas (which are known to be G_{D3} positive) were found to also immunostain strongly for the SGR37 antigen (9). Indeed, in some samples, reactivity with the R24 antibody was found to be more focal, compared to that with SGR37. This implies that naturally occurring tumors *in situ* may have higher levels of de-*N*-acetylation than that of the corresponding cultured cells. Immunoreactivity with SGR37 was also seen in some malignant-appearing cells in lymphoma samples (particularly of the T cell type). Unlike the case with melanomas and lymphomas, there was no SGR37 immunoreactivity in a number of carcinomas studied (R24 immunostained stromal elements in these tumors) (9).

De-*N*-Acetyl Gangliosides Have an Unusual Subcellular Localization.

We used the IgG3 antibody SGR37, and isotype-matched antibodies against G_{D3} (R24) and 9-*O*-acetyl- G_{D3} (27A) to study the distribution of these antigens in cultured Melur and M21 human melanoma cells.(9) Flow cytometric analyses indicated that while G_{D3} and 9-*O*-acetyl- G_{D3} are easily detected on the cell surface, SGR37 reactivity is very low. Glycolipid epitopes could be cryptic because of shielding by cell surface glycoproteins (105) or possibly because of

interactions with other lipids (106). To evaluate Melur cells were treated with either trypsin or saponin. Saponin was used at a concentration of trypsin known to cleave face glycoproteins, exposing underlying glyco probes (105). This treatment did not increase SGR37 reactivity on the cell surface. However, saponin, a reagent commonly used to permeabilize intact cells, clearly increased reactivity to SGR37. In contrast, R24 reactivity on melanoma cells was somewhat increased after trypsin treatment and somewhat decreased after the saponin treatment (9).

Some gangliosides are thought to be selectively located in an intracellular compartment (107-111). Indirect immunofluorescence studies of the saponin-permeabilized cells showed that while the melanoma cells displayed surface amounts of cell surface G_{D3} and 9-*O*-acetyl- G_{D3} , SGR37 reactivity in some internal vesicle-like elements. In contrast, the SGR37 antigen was found to be distributed in a cytoplasmic, and concentrated in the perinuclear region of some cells, with very low levels seen on the cell surface. Saponin is known to permeabilize cells specifically by disrupting cholesterol in cell membranes. Thus, we concluded that SGR37 antigen is either completely intracellular and/or is partly cryptic on the cell surface, in a cholesterol-rich domain which can be disrupted by saponin. Permeabilization of cells with streptolysin O (which makes holes only in the plasma membrane, without increasing cholesterol content) revealed only a small amount of the SGR37 antigen. Thus, the intracellular pool cannot be accessed easily without a membrane-perturbing reagent such as saponin. As an independent check, we also used the MAbs SMR36 (anti de-*N*-acetyl- $G_{M3/D3}$), GMR14 (anti-9-*O*-acetyl- G_{D3}), and Jones (anti-9-*O*-acetyl- G_{D3}). Again, we observed the same differential distribution of these gangliosides. Since both saponin and streptolysin O perturb the plasma membrane in different ways, we also carried out immunofluorescence studies on semi-thin sections of the cultured cells that were harvested without permeabilization (sectioning of the cells was directly done without disrupting membrane structures). This approach strongly corroborated the above results, *i.e.* G_{D3} and 9-*O*-acetyl- G_{D3} are predominantly on the cell surface, whereas de-*N*-acetyl- G_{D3} is detected in a diffuse cytoplasmic internal distribution. The overall conclusion is that the intracellular distribution of the SGR37 antigen is markedly different from those of the other two clo-

related structures(9).

Unlike GD3, de-N-acetyl-GD3 antigen is not enriched in plasma membranes or lysosomes.

Double immunofluorescence studies were performed on the semi-thin sections of cultured cells using the same set of antibodies. R24 and 27A stained the cell surface as well as some intracytoplasmic vesicles which were identified as lysosomes (by the lamp-1 lysosomal marker). In contrast, the diffuse cytoplasmic internal distribution of the SGR37 antigen did not overlap with lamp-1, nor with Calnexin (an ER marker). The signal also did not have a perinuclear Golgi-like pattern. Immunoelectron microscopy with gold-bead tagged antibodies confirmed that the SGR37 antigen was simply scattered in the cytoplasm with no labelling of specific membranous structures. All membranous structures that labelled with R24 were completely devoid of label with SGR-37. Taken together, all of the data showed that the R24 and 27A antigens behaved as expected for typical gangliosides, being located mainly on the plasma membrane and in internal organelles. In contrast, the SGR37 antigen appears to be mainly in the cytoplasm, without any clear association with a membrane-bound organelle.

SGR37 antigen is transiently expressed on the cell surface and then delivered to an internal compartment. De-NAc-G_{D3} is very likely to be derived from G_{D3} by a de-N-acetylation reaction, after the initial synthesis of G_{D3} in the Golgi (6,8). Thus, we can ask if the De-NAc variant is targeted directly to its internal compartment from the Golgi, or whether it first passes through the plasma membrane. To study this, we exploited the fact that when antibodies to potential cell surface antigens are added to the medium of live cells, adsorptive endocytosis is antigen-dependent, and the final intracellular distribution of the antibody probe indicates the distribution of the antigen (112–115). The melanoma cells were incubated with the different monoclonal antibodies under standard culture conditions, and uptake of antibodies monitored. Although it was possible to observe some antibody internalization most of mAb R24 was retained on the cell surface. On the other hand, SGR37 was internalized and concentrated in cytoplasmic vesicular structures. Previous studies have indicated that internalized gangliosides are typically delivered to lysosomes (78,116). In order to test this, cells were simultaneously pulsed with mAb SGR37 along with dextran-FITC, which is known to be con-

centrated in lysosomes. We found that SGR37 and dextran-FITC were concentrated into different compartments. Thus, the SGR37 antigen is at least transiently concentrated on the cell surface, but then becomes predominantly intracellular, concentrating initially in an unknown location other than in functional lysosomes.

SGR37 antigen accumulates on the surface of non-adherent cells, but this is not related to cell cycle status.

In studies of cultured melanoma cells, we found that SGR37 reactivity was higher in the small rounded-up cells, as compared to the bulk population of adherent cells. Flow cytometry confirmed that the cells had an increased cell surface reactivity to SGR37, but not to R24. This fit with our previous results that microtubule-disrupting agent nocodazole increases the surface density of SGR37 antigen (8). The proportion of rounded-up cells in different cell cycle phases varied, but was not significantly different from the bulk population. No significant variation in either cell cycle phase or global expression of SGR37 antigen was observed. SGR37 antigen accumulates on the surface of non-adherent or rounded-up cells regardless of the cell cycle phase.

In future, it will be very interesting to explore the marked differential subcellular localization of SGR37 antigen, which differs from G_{D3} solely by the presence of a free amino group (the absence of the N-acetyl group). In this regard, our data suggest that De-N-acetyl-G_{D3} is not simply a terminal degradation product of G_{D3}, but well with our early studies where we had noted the presence of a re-N-acetylase activity, which could be derived from endogenous De-NAc-G_{D3} (6). The functional consequences of De-NAc-G_{D3} expression also need to be explored. Others have also shown that exogenously added gangliosides can block adhesion of cells to extracellular matrix (117). In other work described in this volume, replacement of the sialic acid *N*-acetyl group by *N*-propionyl or other acyl chains achieved on cells with mannosamine analogs led to loss of growth control and loss of contact-inhibition (118). The effects of *in vitro* addition of G_{M3} (growth suppressor) or de-N-acetyl-GM3 (growth enhancement) to cells (5,7) also provides a framework for future studies. The discovery by Kannagi and colleague

neuraminic acids can be converted into intramolecular lactams (described elsewhere in this volume) adds further complexity to the study of de-*N*-acetyl-gangliosides. Many other questions regarding the biosynthesis, trafficking and function of these novel gangliosides remain to be studied. Meanwhile, the low levels of the SGR37 antigen in normal tissues, the high levels in some tumors, its selective endocytosis from the cell surface, and its high expression on rounded-up/non-adherent cells suggests that it is worthwhile to explore diagnostic or therapeutic approaches using this antibody.

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